

Research article

Long COVID, Vaccine Injury and Gut Dysbiosis

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Abstract

Introduction. COVID-19 infection can trigger a chronic condition known as Long COVID, which as of today, still affects millions of U.S. adults and children and remains a major public health concern. In this study, we investigated whether Long COVID patients have a distinct gut microbiome profile when compared to healthy controls. Similarly, with the prevalence of patients reporting Long COVID symptoms upon vaccination for SARS-CoV-2, we also compared the gut microbiome profiles of these patients with those of healthy controls.

Methods. Fecal samples from 31 long COVID patients, 37 COVID-19 Vaccine-Injured, and 20 healthy controls were collected at ProgenaBiome laboratories. Microbiota composition was compared using shotgun methodology and next-generation sequencing (NGS) on the Illumina NextSeq with 500/550 High-Output Kits V.2.5 for 300 cycles. Data were processed through the OneCodex bioinformatics system to determine microbiota composition and relative abundances of top genera and phyla. A Mann-Whitney U test was used to evaluate statistical significance.

Results. We found that *Bacteroides* was the dominating microbe in both Long COVID and SARS-CoV-2 Vaccine-Injured patients. Compared to healthy controls, Long COVID patients' gut microbiome had more *Bacteroides* ($p < 0.001$) and less *Bifidobacterium* ($p < 0.001$) and *Faecalibacterium* ($p = 0.002$). SARS-CoV-2 vaccine-injured patients had less *Bifidobacterium* ($p < 0.001$) and *Faecalibacterium* ($p = 0.002$), compared to healthy controls.

Conclusions. These data suggest that gut dysbiosis may be associated with Long COVID and SARS-CoV-2 Vaccine-Injury. Future research is needed to confirm these findings. However, if confirmed, Long COVID and Vaccine-Injured patients may be treated for dysbiosis to alleviate symptoms.

Keywords: SARS-CoV-2, Long COVID, Vaccine-Injured, NGS, microbiome

Introduction

Since the onset of SARS-CoV-2 in December 2019 as a global pandemic, many individuals have developed chronic conditions after exposure to the SARS-CoV-2 virus or as a reaction to SARS-CoV-2 vaccination. According to the Household Pulse Survey, 29.8% of adults in the U.S. have had or currently have post-COVID symptoms lasting longer than 3 months [1]. Often these symptoms include chronic fatigue syndrome (ME/CFS), muscle weakness, and diarrhea [2]. Understanding why these patients experience chronic symptoms has become paramount in

researchers' attempts to understand the full breadth of COVID-19's impact on patient wellbeing that goes well beyond the acute viral infection. As SARS-CoV-2 likely induces immune dysregulation through mechanisms like cytokine storms and mitochondrial oxidative stress, focusing on the host's immune response would be one way to address Long COVID [3,4]. As the body's largest housing for its immune response, studying dysregulation of the gastrointestinal tract is a logical focus for elucidating issues of complications after host infection [3].

Beyond SARS-CoV-2, it has been shown that many chronic

inflammatory disorders such as autoimmune diseases, allergies, and metabolic syndromes, are correlated to a deteriorated symbiosis of gut microbes in the intestine [2,5]. For SARS-CoV-2, there is a higher risk for severe infection and chronic fatigue when the virus invades patients with comorbidities like chronic obstructive pulmonary disease (COPD), which could be linked to a weak gut microbiota [6,7]. In more severe cases of SARS-CoV-2, intestinal conditions such as inflammatory bowel disease (IBD) show higher prevalence, existing in approximately 17.6% of all cases [3,8].

In successfully recovered COVID-19 patients, higher Bifidobacterium and Faecalibacterium abundances have been correlated with increased spike IgG titers [9]. As Bifidobacterium abundance is known to diminish due to certain illnesses, including viral infections like influenza, it is anticipated that a similar trend specifically linked to Bifidobacterium would be of some cause for severity and chronicity of symptoms during and after SARS-CoV-2 infection [2,10]. It is, indeed, observed that COVID-19 patients experiencing hair loss have previously shown to have reduced Bifidobacterium and Faecalibacterium [11].

It has been recently shown that SARS-CoV-2 vaccinated individuals also experience chronic symptoms similar to those observed in Long-COVID [5,12-19]. Indeed, Bhattacharjee et al., 2025, in a cross-sectional study among 42 post-SARS-CoV-2 vaccine syndrome participants and 22 healthy controls enrolled in the Yale LISTEN study, found that the SARS-CoV-2 vaccinated participants had diminished immune function markers compared to controls [12].

To evaluate whether Long-COVID and SARS-CoV-2 Vaccine-Injury patients share a pattern in gut microbiota changes compared to healthy controls, we measured gut microbiome abundance and composition among these groups.

Methods

Study subjects who attended a GI clinic were recruited for this study. Inclusion criteria were no symptoms reported (controls),

the presence of GI symptoms and/or Long COVID/Vaccine-Injured symptoms, and aged 18 yrs. and older. Participants were excluded if they were younger than 18 yrs. old and if symptoms were severe enough to require hospitalization. A total of (n = 88) consecutive participants who attended the GI and gave written informed consent to participate in this study were included. Fecal samples from 31 Long COVID patients, 37 COVID-19 Vaccine-Injured, and 20 healthy controls were collected at ProgenaBiome Laboratories.

Microbiota composition was compared using DNA quantification, purification, and normalization using shotgun methodology and next-generation sequencing (NGS) on the Illumina NextSeq with 500/550 High-Output Kits V.2.5 for 300 cycles. Data were processed through the OneCodex bioinformatics system to determine microbiota composition and relative abundances of top genera and phyla. A Mann-Whitney U test was used to evaluate statistical significance.

Results

Patient characteristics

The demographics of all patients from the Control (n=20), Long COVID (n=31), and Vaccine-Injured (n=37) groups are presented in Table 1. Among the controls, the average age and standard error of the mean was 46±4 years; 10 of 20 (50%) were male; 18 out of 20 (90%) self-identified as non-Hispanic White, 1 (5%) identified as Asian, and 1 as Native American (5%).

Among the Long COVID patients, the average age and standard error of the mean was 50±2.7 years; 12 of the 31 (39%) were male; 29 out of 31 (94%) self-identified as White, 1 patient (3%) identified as Asian, and one (3%) as Hispanic. The average age and standard error of the mean, for the Vaccine-Injured group was 51±2.6 years; 21 of the 37 (57%) were male; 33 of the 37 (89%) self-identified as non-Hispanic White, 1 (3%) identified as black, 1 (3%) identified as Asian, and 2 (5%) identified as Hispanic. Both Long COVID and Vaccine-Injured patients had more than one underlying comorbidity considered risk factors,

Table 1. Demographics of study participants

	Control (n=20)	Vaccine Injured (n=37)	Long COVID (n= 31)
Age (mean, SEM, range)	46, 18, 20-87	51, 2.6, 16-81	50, 2.7, 19-85
Male (n, %)	10 (50%)	21 (57%)	12 (39%)
Female (n, %)	10 (50%)	16 (43%)	19 (61%)
Race (n, %)	# Of Subjects (%)	# Of Subjects (%)	# Of Subjects (%)
White	18 (90%)	33 (89%)	29 (94%)
Black	0 (0%)	1 (3%)	0 (0%)
Asian	1 (5%)	1(3%)	1 (3%)
Hispanic	0 (0%)	2 (5%)	1 (3%)
Native American	1 (5%)	0 (0%)	0 (0%)
# Of Comorbidities	# Of Subjects (%)	# Of Subjects (%)	# Of Subjects (%)
0	0 (0%)	0 (0%)	0 (0%)
1	0 (0%)	21 (57%)	20 (65%)
2	0 (0%)	14 (38%)	9 (29%)
3	0 (0%)	2 (5%)	1 (3%)
4	0 (0%)	0 (0%)	1 (3%)

including obesity, cardiovascular conditions, and being immunocompromised.

Microbiome diversity and composition

NextGen Sequencing analysis revealed that, at the genus level, shown in Figure 1, Bacteroides are the dominating microbe in both Long COVID and Vaccine-Injured patients. Compared to healthy controls, Long COVID patients' gut microbiome had more Bacteroides ($p < 0.001$) and Blautia ($p = 0.039$), while having less Bifidobacterium ($p < 0.001$), Faecalibacterium ($p = 0.002$), and Collinsella ($p = 0.004$). Vaccine-Injured patients had elevated Bacteroides ($p = 0.168$), and less Bifidobacterium ($p < 0.001$), Faecalibacterium ($p = 0.002$), Collinsella ($p = 0.002$), and Ruminococcus ($p = 0.034$) compared to healthy controls, Figure 1. All

other species did not indicate any significant difference between patient categories.

At the Phylum level, Figure 2, both Long COVID and Vaccine-Injured patients had less Actinomycetota and Bacillota, while more abundance of Bacteroidota and Firmicutes, compared to controls, all $p < 0.05$.

Long COVID patients' gut microbiome had less overall diversity, indicated by the Shannon Index, compared to controls, Figure 3, however, the association was not statistically significant ($p = 0.065$). Likewise, the Shannon Index for diversity was not statistically significant when comparing Vaccine-Injured gut microbiome with controls, Figure 4.

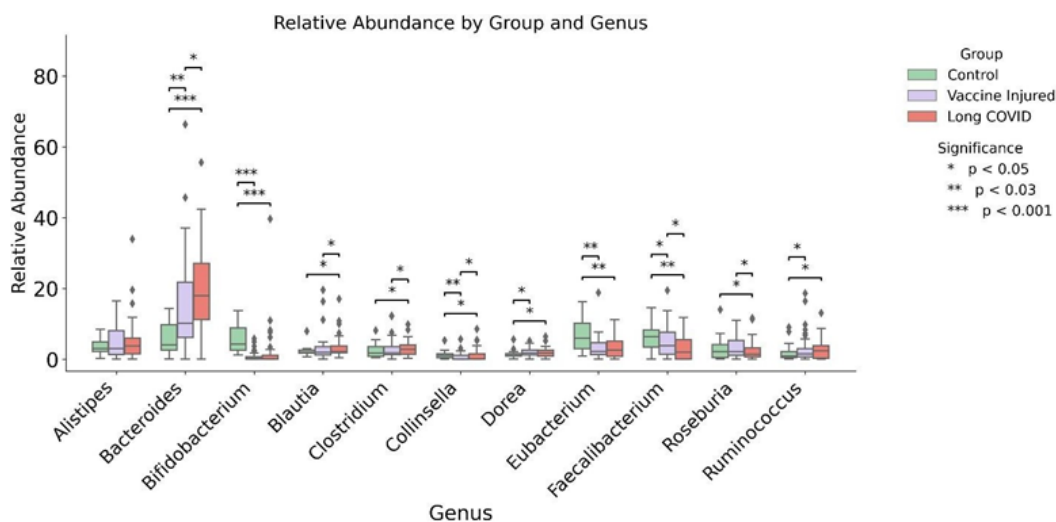


Figure 1. Controls versus Vaccine Injured and Long COVID at Genus level

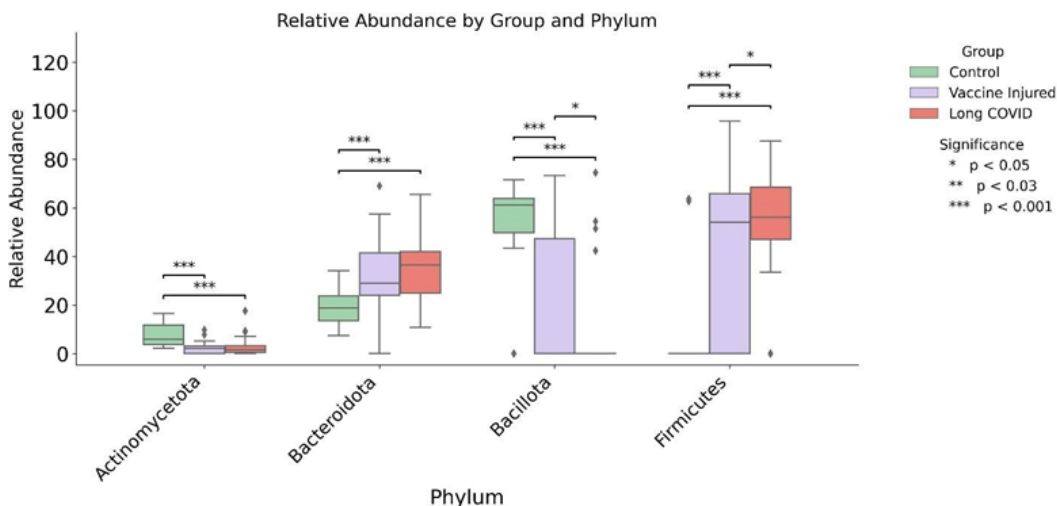


Figure 2. Controls versus Vaccine Injured and Long COVID at Phylum level

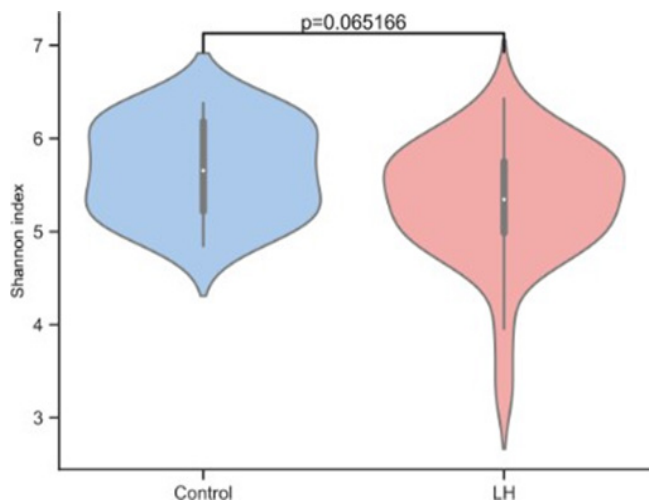


Figure 3. Gut Microbiome Diversity in Long COVID patients versus controls

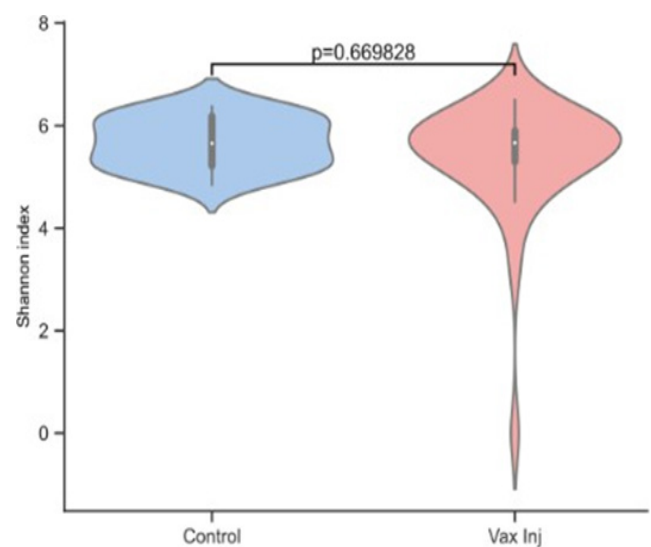


Figure 4. Gut Microbiome Diversity in Vaccine-Injured patients versus controls

Discussion

This study found that both Long COVID patients and SARS-CoV-2 vaccinated individuals had elevated levels of gut Bacteroides and decreased levels of gut Bifidobacterium and Faecalibacterium. If confirmed in larger studies, manipulating gut microbiota may serve as a concomitant treatment for Long COVID therapies.

Across Long COVID and post-SARS-CoV-2 vaccination syndrome cases, clinical and translational studies show characteristic, and often persistent, gut dysbiosis that tracks with disease severity and long-term symptoms. Indeed, Gu et al. [10] in a study among 30 COVID-19 patients, 24 H1N1 influenza, and 30 controls, reported significantly reduced bacterial diversity in COVID-19 patients, enrichment of genera such as Streptococcus, Rothia, Veillonella, and Actinomyces, and depletion of “beneficial symbionts” while controls had higher prevalence of beneficial Bifidobacterium, Blautia, Romboutsia, and Collinsella. Using a five-taxon signature discriminated COVID-19 from controls with 95% confidence interval, and seven taxa distinguished COVID-19 from H1N1 also with a 95% confidence interval [10].

In a qPCR-based study of 57 patients with general, severe, or critical COVID-19 disease, Tang et al. [20] quantified large reductions in key butyrate producers across the spectrum of severity. For example, the median Bifidobacterium abundance fell from 5.7×10^6 common logarithm of copy number per gram in the general group to 4.4×10^6 in the severe group, and 1.4×10^4 in the critical group with similar stepwise declines in *F. prausnitzii*, *Clostridium leptum*, *C. butyricum*, and *Eubacterium rectale*, while *Enterococcus* and *Enterobacteriaceae* expanded [20]. Notably, the Ec/E ratio was able to predict mortality in critically ill patients.

Longitudinal metagenomics in smaller cohorts linked this dysbiosis to intestinal viral activity. Zuo et al. [21] followed 15 hospitalized COVID-19 patients and found that 47% of them had faecal SARS-CoV-2 by RNA metagenomics, with a “high infectivity” signature (increased 3′ - 5′ end coverage) persisting up to 6 days after respiratory PCR negativity; stools with this signature were enriched in *Collinsella aerofaciens*, *Collinsella tanakaei*, *Streptococcus infantis*, and *Morganella morganii*, whereas

samples with low-to-none faecal viral activity were enriched in SCFA producers including *Parabacteroides merdae*, *Bacteroides stercoris*, *Alistipes onderdonkii*, and *Lachnospiraceae bacterium 1_1_57FAA* [21].

Larger shotgun-based cohorts extend these associations into the post-acute phase. In a prospective study of 106 patients followed for 6 months with 68 non-COVID controls, Liu et al. found that 76% met criteria for post-acute COVID-19 syndrome and at 6 months, these patients remained enriched for *Ruminococcus gnavus* and *Bacteroides vulgatus* with depleted *F. prausnitzii* and *Bifidobacterium pseudocatenulatum* [11].

A complementary multi-kingdom analysis of 133 patients, 296 fecal metagenomes and 79 fecal metabolomes, defined two reproducible gut configurations. One was characterized as having a low-diversity, pathogen-rich cluster associated with more severe acute disease, systemic inflammation, and a higher prevalence of post-acute COVID-19 syndrome at 6 months, and a second cluster linked to milder illness and faster recovery [22].

Importantly, gut microbiome and SARS-CoV-2 vaccine interactions appear to differ in individuals with pre-existing immunocompromised conditions. In a cohort of 143 individuals involving people living with HIV and receiving BNT162b2, Ray et al. [9] showed that baseline microbiota composition and diversity predicted vaccine immunogenicity. High spike IgG and CD4⁺ T-cell responders exhibited lower α -diversity but relative enrichment of Bifidobacterium and Faecalibacterium, whereas *Cloacibacillus* was associated with poor antibody responses- suggesting that pre-existing microbiome configurations can modulate both the magnitude and quality of vaccine-induced immunity [9]. With evidence that specific gut bacteria influence both susceptibility to and outcomes of SARS-CoV-2 infection, these findings support a model in which SARS-CoV-2 illness, Long COVID, and post-vaccination syndromes arise on a background of gut microbial dysregulation and, in turn, further perpetuate this dysbiosis.

These microbial shifts may influence immune regulation that contributes to the wide range of persistent symptoms. Possible

mechanisms have been identified in earlier studies which may underly the myriad of both Long COVID and vaccine-related adverse events but have not been definitively linked to the microbiota. Marked by cytokine storms, COVID-19 infection reflects a profound immune overactivation [24]. Studies have highlighted a persistent small subset of patients that do not respond positively upon vaccination, sometimes reporting symptoms within a day after vaccination [22]. In patients experiencing post-vaccination syndromes (PVS), immunological imbalances have been reported with specific signatures compared to healthy controls, including reduced circulating memory and effector T cells, and increased TNF α -producing CD8+ T cells along with an increased likelihood of Epstein-Barr virus reactivation [12]. A strong correlation was shown between the number of vaccine doses and plasma anti-S IgG. In cases of vaccine-induced immune thrombotic thrombocytopenia (VITT), immune responses have been shown to trigger the production of anti-platelet factor 4 (PF4) antibodies [17]. Post-vaccination autoimmune myocarditis has been linked to CD4+ T cell-mediated, with elevated levels of IL-1 α and IL-1 β , cytokines central to the vaccine immune response, especially for male patients that suggests a sex hormone influence [18].

Neurological complications, including Guillain-Barré syndrome for patients with a predisposition, have been observed, likely due to the production of spike proteins that leads to autoimmune demyelination [17]. Moreover, free circulating spike protein has been detected exclusively in symptomatic individuals and has been correlated with cardiac troponin T, indicating a possible role in cardiac inflammation and injury [18].

The vast diversity of bacteria in the human body suggests a range of natural mechanisms in immune regulation, many of which remain poorly understood [25]. Some bacteria actively participate in the body's regulation of the immune system, acting through a pro-inflammatory means while others work through an anti-inflammatory mechanism [8]. As key components of immune regulation depend on microbial metabolites, disruptions in the microbiome may influence both susceptibility to infection and disease severity [5].

As the primary target for active SARS-CoV-2, angiotensin-converting enzyme 2 (ACE2) receptors indicate likely the first effected cellular mechanism and the first connection the gut microbiome has to bodily infection. The intestine is known to express a higher density of ACE2 receptors than most other tissue [19,22]. While ACE2 allows the entry of SARS-CoV-2 into the body through the intestine, it allows barrier function impairment, including nutrient absorption, while acting as a regulator for the gut microbiota [10,13,14].

Short-chain fatty acids (SCFAs) must also be considered. SCFAs are utilized in the production of O-glycans that aid in the permeability of the intestinal wall for translocation of pathogens and toxins [14,26]. The permeability of the intestinal wall is affected by SCFAs, L-isoleucine, and bile acids. Losing the production of SCFAs through loss of Bifidobacterium, Ruminococcus, and Clostridium also elevate the inflammatory response by disrupting the adaptive immune system by not regulating T-cell development through GPCR. With a reduction in SCFAs, as well as L-isoleucine, the largest immune reservoir housed in the

intestine is weakened [27]. Patients with chronic inflammatory symptoms will also express higher levels of pro-inflammatory cytokines IL-18, IL-1 β , and IL-6, which increase lactic acid production that can lead to further pain [28,29].

Correlated to gut microbiota health, higher IgG titers in some individuals were associated with greater abundance of Bifidobacterium and Faecalibacterium, though paradoxically accompanied by reduced alpha diversity [9]. Along with Faecalibacterium prausnitzii, Roseburia and Ruminococcus, are butyrate producing bacteria whose reduced abundances have also been correlated with negative outcomes of SARS-CoV-2 infection, including hair loss [11].

While most changes in the microbiome are decreases in bacterial abundances, the biggest change appears in the increase of Bacteroides, which have been shown to exacerbate pro-inflammatory responses in the intestine through the release of TNF- α and immune cells such as IFN- γ regulatory CD4+ T cells [5,30,31]. A correlation is seen in the overabundance of Bacteroides with the overexpression of lectin reg III beta fusion protein which is already known to be involved in most states of inflammation in the intestine [32,33].

With the goal of treating conditions such as inflammatory bowel disease through modulation of the gut microbiome, interventions including probiotic supplementation and fecal microbiota transplantation (FMT) have shown therapeutic benefit. These approaches have been associated with increased relative abundances of key beneficial taxa, particularly

Bifidobacterium and Faecalibacterium [34,35]. If severity of SARS-CoV-2 infection does present a correlative linkage to the gut microbiome, similar treatments would be beneficial for patients experiencing chronic symptoms post-infection or post-vaccination.

Study limitations include the relatively small sample size, and analyses were not adjusted for confounders. However, the observed gut microbiota alterations were distinguishable across patient groups.

Conclusions

While acute SARS-CoV-2 infection was initially the concern, attention has shifted toward the growing number of individuals experiencing long-term effects following infection or vaccination. Cases of Long COVID and adverse reactions to SARS-CoV-2 vaccines can persist for years, with symptoms including, but not limited to, muscle pain, fatigue, shortness of breath, and anxiety. Many individuals are forced to adjust their lifestyles to accommodate the ongoing nature of chronic illness, often without knowing when or if symptoms will subside. This uncertainty highlights an urgent need for effective and lasting therapeutic strategies. In an effort to explore the role of microbial dysbiosis in these persistent conditions, our findings reveal consistent alterations in the gut microbiota, which are common among affected individuals.

With targeted interventions such as probiotic treatment, modulation of the gut microbiome may offer a gradual path to recovery and an adjunctive approach to Long COVID and vaccine injury management.

Declarations

Ethics approval and consent to participate

This study was approved by the institutional review board of IRCM IRB. Consent to participate in this study from each patient was agreed via signed informed consent.

Consent for publication

Agreed to by patients on the signed informed consent, demonstrating that the patient understood the procedures and the purpose of the study.

Availability of data and materials

The datasets used and/or analyzed during the current study are available from the corresponding author on reasonable request.

Competing interests

Dr. Hazan is the CEO of Progenabiome and the owner of Microbiome Research Foundation.

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Authors' contributions

Conceptualization, BR and SH; Methodology, BR, AG, and SH; Investigation, AV and SH. Writing-original draft, BR; Writing-review and editing, BR, AV, and SH Supervision, SH; Funding acquisition, SH. All authors read and approved the final manuscript.

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